

Volatilization of Mercury Compounds by Methylmercury— Volatilizing Bacteria in Minamata Bay Sediment

Kunihiko Nakamura, ¹ Taizo Sakata, ² and Hideomi Nakahara³

¹Department of Basic Medical Science, National Institute for Minamata Disease, 4058-18 Hama, Minamata City Kumamoto 867, Japan, ²Laboratory of Microbiology, Faculty of Fisheries, Kagoshima University, Kagoshima 890, Japan, and ³Department of Environmental Health, Medical University of Yamanashi, Yamanashi 400-38, Japan

Minamata Bay has been heavily polluted by high mercury concentrations which gave rise for a long time to methylmercury poisoning, Minamata disease (Kutsuna 1968; Irukayama 1977). The mercury still exists in the sediments of the Bay (Kumamoto Prefecture Government 1973; Kumagai and Nishimura 1978). The fish and the shellfish living in the Bay contain high methylmercury concentrations as compared with those in other marine environments. The population of mercury-resistant bacteria in the sediments of Minamata Bay is larger than that in the sediments of other marine environments (Nakamura $et\,al.$ 1986).

The mercury-resistant bacteria isolated from a marine environment have been found to transform organic and inorganic mercury compounds into mercury vapor ($\rm Hg^o$) (Nelson et~al.~1973; Spangler et~al.~1973; Olsen et~al.~1979). The mercury-resistance confirmed in various bacterial genera has been shown to be plasmid-mediated volatilization (Clark et~al.~1977; Weiss et~al.~1977; Robinson and Tuovinen 1984). However, there has been little definitive information on the volatilization of organic mercury by the bacteria living in the mercury-polluted environment.

It is important to know what bacterial transformations of mercury have been taking place and how the mercury-resistant bacteria may be playing a role in the mercury cycle in the marine environment of Minamata Bay. The object of the present study is to clarify the characteristics of the methylmercury-volatilizing bacteria in the sediments of Minamata Bay and of the volatilization of various mercury compounds by these bacteria.

MATERIALS AND METHODS

Sediment samples were collected in October 1985 from two stations in Minamata Bay near the drainage outlet of the acetoaldehyde plant of Minamata Factory by using sterile cylinders (55 cm diameter). The topmost 3 cm of the sediments were used for assessing total mercury contents and for isolating the bacteria.

Send reprint requests to K.Nakamura at the above address.

The basal medium was artificial sea water medium (modified ZoBell 2216 E medium) as described previously (Nakamura $et\ al.$ 1986). About 1000 strains of methylmercury-resistant bacteria were isolated on agar plates containing 0.1 µg/mL of CH3HgCl after incubation at 25°C for 48 h. These isolates were screened for the ability to volatilize methylmercury by measuring mercury loss in liquid media containing methylmercury (0.1 µg/mL) after incubation for 48 h at 25°C. The total mercury content in the medium was analyzed by a flameless atomic adsorption spectrophotometer with mercury analysis vaporizer (Tanaka $et\ al.$ 1974).

The methylmercury-volatilizing strains were classified accordance with the taxonomic scheme proposed by Shewan et al. (1960) and "Bergye's Manual of Determinative Bacteriology" (Buchanan and Gibbons 1974). The minimal inhibitory concentrations (MICs) of mercury chloride (HgCl₂), methylmercury chloride (CH₂HgCl), ethylmercury chloride (C₂H₅HgCl), propylmercury chloride (C₃H₇HgCl), and phenylmercuric acetate (C6H5HgOCOCH3) against the methylmercury-volatilizing strains were determined by the agar dilution method. MICs of mercury compounds were taken as the next concentration beyond the last concentration at which growth was observed on solid medium in the presence of The mercury sequential concentrations of mercury compounds. compounds and their concentrations ($\mu g/mL$) used for the volatilization were as follows: HgCl2, 5; CH2HgCl, 0.25; $\rm C_2H_5HgC1,\ 0.1$; $\rm C_3H_7HgC1,\ 0.1$; $\rm C_6H_5Hg0C0CH_3,\ 1.$ The mercury contents in the sediments were directly analyzed by the same method as used in analyzing the total mercury content in the medium.

RESULTS AND DISCUSSION

The total mercury concentration and the percentage of methylmercury-resistant bacteria for the total bacterial counts in the sediment of Minamata Bay are as shown in Table 1. The percentage of methylmercury-resistant bacteria for the total bacterial counts was calculated from the separate counts on methylmercury-free agar plates. The total mercury concentrations in the sediment of two stations in Minamata Bay were 32.4 $\mu g/mL$ and 23.5 $\mu g/mL$, respectively. One observation made was the tendency for the percentage of methylmercury-resistant bacteria to be higher where the mercury concentration in the sediment was also higher.; a similar tendency was reported by Nakamura $et\,al.$ (1986).

The percentage and generic composition of the methylmercury-volatilizing bacteria are shown in Table 2. A total of 1068 bacterial strains (965 non-swarmer and 103 swarmer) isolated from the agar plates containing 0.1 μ g/mL methylmercury were screened for methylmercury volatilizing ability. Of these strains, only 80 strains (7.5%) found to volatilize methylmercury by measuring mercury loss in liquid media containing methylmercury. These strains were composed of 60 Bacillus species (75.0%), 15 Pseudomonas sp. (18.8%), 3 Moraxella sp. (3.7%), and 2 unclassified (2.5%). The two unclassified strains were gram-

Table 1. Total mercury concentrations and percentage of CH₃HgCl-resistant bacteria in the sediments of Minamata Bay.

	Mercury ^a (μg/g)	C.F.U. ^b	CH ₃ HgC1		(µg/mL)	
	(1 3. 37		0.01	0.025	0.05	0.1
Station 1	32.4	2.2×10 ³	81.8c	63.6	54.5	2.3
Station 2	23.5	6.0×10 ³	76.6	48.3	36.7	3.7

a Total mercury concentrations in sediment (/ dry weight).

bColony-forming units.

Table 2. Generic composition of CH₃HgC1-volatilizing bacteria in the sediments of Minamata Bay.

No. of tested strains	No. of CH ₃ HgCl- volatilizing bacteria	Bacterial genera		
1068 (non-swarmer-965) (swarmer103)	80 (7.5 %)	Bacillus 60 (75.0 %) Pseudomonas 15 (18.8 %) Moraxella 3 (3.7 %) unclassified 2 (2.5 %)		

negative rods with peritrichous flagella. The distribution rate of MICs for various mercury compounds determined on 80 methylmercury-volatilizing strains from Minamata Bay sediments is shown in Table 3. The MICs of HgCl $_2$ and C $_6$ H $_5$ HgOCOCH $_3$ against these strains are distributed in narrow ranges: HgCl $_2$, 80-160 μ g/mL; C $_6$ H $_5$ HgOCOCH $_3$, 4-8 μ g/mL. On the other hand, those of CH $_3$ HgCl, C $_2$ H $_5$ HgCl and C $_3$ H $_7$ HgCl are widely distributed. It is hypothesized that the differences depend on the solubility of mercury compounds in lipids in bacterial membranes as well as on the binding of the compounds to sulfhydryl groups of proteins and enzymes, as reported by Berlin (1979).

The volatilization of mercury compounds by the methylmercury-volatilizing bacteria is shown in Table 4. All of the methylmercury-volatilizing bacteria were able to volatilize each of the mercury compounds used in this study. The mercury resistant plasmid of $Pseudomonas\ aeruginosa$ and $Escherichia\ coli$ is

Table 3. Distribution of Minimal Inhibitory Concentration (MICs) of mercury compounds against CH₃HgCl-volatilizing bacteria.

	MIC (µg/ml)	$Bacillus$ $(60)^a$	Pseudo- monas (15)	Mora- xella (3)	Unclassi- fied (2)
	40	0.0 ^b	6.7	0.0	0.0
HgC1 ₂	80	11.7	0.0	0.0	0.0
119012	160	83.4	93.3	100.0	100.0
	320	5.0	0.0	0.0	0.0
	0.25	13.3	21.0	33.3	0.0
	0.5	35.0	46.7	33.3	100.0
CH ₃ HgC1	1	26.7	20.0	33.3	0.0
3 0	2	23.3	13.3	0.0	0.0
	4	1.7	0.0	0.0	0.0
	0.25	3.3	6.7	0.0	0.0
C ₂ H ₅ HgC1	0.5	38.4	53.3	0.0	50.0
2 3	1	26.7	6.7	100.0	50.0
	2	31.6	33.3	0.0	0.0
	0.25	5.0	20.0	0.0	0.0
	0.5	41.7	33.3	0.0	100.0
C ₃ H ₇ HgC1	1	20.0	20.0	0.0	0.0
3 / 3	2	20.0	13.4	75.0	0.0
	4	1.3	13.3	25.0	0.0
	0.5	0.0	0.0	0.0	50.0
C ₆ H ₅ HgOCOCH ₃	2	16.7	26.7	0.0	0.0
0.5 - 5	4	5.0	6.8	0.0	0.0
	8	78.3	66.7	100.0	50.0

^aNumber of strains ^bPercentages of strains

divided into two classes of resistance: (1) "narrow-spectrum" resistance plasmids that are resistant to Hg and (2) "broad-spectrum" resistance to Hg , CH3HgCl, C2H5HgCl, and C3H7HgCl and C6H5Hg0C0CH3 (Clark $et\ al$ and Weiss $et\ al$. 1977). The volatilization of C3H7HgCl by bacteria had not yet been reported. It was confirmed in this study that 80 methylmercury-volatilizing bacteria removed mercury from liquid culture containing C3H7HgCl. It is probable that mercury-resistant bacteria remove C3H7HgCl as well as CH3HgCl and C2H5HgCl through volatilization.

The methylmercury-volatilizing bacteria were isolated on agar plates containing 0.1 μ g/mL of CH₃HgCl and screened for the ability to volatilize methylmercury. The percentages of methylmercury-volatilizing bacteria for the total bacterial count were low as compared to those of HgCl₂-volatilizing bacteria as reported by Nakamura et~al.~(1986). Spangler et~al.~(1973) reported

Table 4. Volatilization (%) of mercury compounds by CH₃HgCl-volatilizing bacteria isolated from Minamata Bay sediments.

	Bacillus (60)ª	Pseudomo- nas (15)	Moraxella (3)	Unclassi- fied (2)
HgC1 ₂	76.8±1.0 ^b	77.5±2.2	73.2±7.8	83.9±1.6
CH ₃ HgC1	88.5±0.3	88.1±0.8	87.0±3.6	89.1±0.0
C ₂ H ₅ HgC1	77.9±0.6	76.6±2.4	81.5±0.9	74.3±2.0
C ₃ H ₇ HgC1	77.9±0.6	73.9±1.5	81.2±0.9	77.8±3.3
C ₆ H ₅ Hg0C0CH ₃	44.6±0.8	47.2±1.2	44.9±3.2	44.6±1.1

aNumber of tested strains

that 30 of 207 (14.5%) bacteria, isolated from the sediment of Lake St Clair, were found to be methylmercury-volatilizing strains. The generic composition of those methylmercury-volatilizing bacteria were similar to that of bacteria in the sediment of Minamata Bay. These results suggest that mercury does not play a large role in causing bacteria in the sediment of Minamata Bay to develop their capacity for methylmercury-resistance through volatilization.

There are many reports concerning methylmercury formation in aquatic environments (Jensen and Jernelöv 1969; Andren and Harris 1973; Olsen and Cooper 1976). However, methylmercury is not detectable in the sediment of Minamata Bay. The methylmercury-volatilizing bacteria may play an important role in the mercury cycle of Minamata Bay.

Acknowledgments. The authors would like to thank the staff of the Pollution Control Office, Kumamoto Prefecture, for help in sampling.

REFERENCES

Andren AW, Harriss RC (1973) Methylmercury in esturine sediments. Nature 245:256-257

Berlin M (1979) Mercury. In: Friberg, Norberg F, Vouk VB (ed) Handbook on the toxicology of metals, Elsevier, Amsterdam, pp 503-530

Buchanan RE, Gibbons RE (1974) "Bergey's Manual of Determinative Bacteriology", 8th ed, Williams & Wilkins, Baltimore, pp 217-351 Clark DL, Weiss AA, Silver S (1977) Mercury and organomercurial resistances determined by plasmids in *Pseudomonas*. J

^bMean loss ± SD (%)

- Bacteriol 132:186-196
- Irukayama K (1977) Case history of Minamata disease. In : Taubaki
 T, Irukayama K (eds.) Minamata disease, Elsevier, New York,
 pp 1-56
- Jensen S, Jernelöv A (1969) Biological methylation of mercury in aquatic organisms. Nature 223:753-754
- Kumagai M, Nishimura H (1978) Mercury distribution in seawater in Minamata Bay and the origin of particulate mercury. J Oceanogr Soc Jap 34:50-56
- Kumamoto Prefecture Government (1973) White paper on environmental pollution (in Japanese), Kumamoto Prefecture, Kumamoto, pp 145-152
- Kutsuna M (1968) Minamata disease. Study group of Minamata disease. Kumamoto University, Kumamoto, pp 1-228
- Nakamura K, Fujisaki T, Tamashiro H (1986) Characteristics of Hgresistant bacteria isolated from Minamata Bay sediment. Environ Res 40:58-67
- Nelson JD, Blair W, Brinckman FE, Colwell RR, Iverson WP (1973) Biodegradation of phenylmercuric acetate by mercuryresistant bacteria. Appl Environ Microbiol 26:321-326
- Olson BH, Barkay T, Colwell RR (1979) Role of plasmid in mercury transformation by bacteria isolated from the aquatic environment. Appl Environ Microbiol 28:478-485
- Olson BH, Cooper RC (1976) Comparison of aerobic and anaerobic methylation of mercuric chloride by San Francisco Bay sediment. Water Res 10:113-116
- Robinson JB, Tuovinen O (1984) Mechanisms of microbial resistace and detoxification of mercury and organomercury compounds: Physiological, biochemical, and genetic analyses. Microbiol Rev 48:95-124
- Shewan JM, Hofbbs G, Hogkinss W (1969) A determinative scheme for the identification of certain genera of Gram-negative bacteria, with special reference to the *Pseudomonasdacea*. J Appl Bacteriol 23:379-390
- Spangler WJ, Spigarelli JL, Rose JM, Flippin RS, Miller HH (1973) Degradation of methylmercury by bacteria isolated from environmental samples. Appl Microbiol 25:488-493
- Tanaka K, Fukaya K, Fukai S, Sugano S (1974) Hygienic chemical studies on mercury III. Rapid flameless atomic absorpiometric determination of organic mercury by quartz tube combustion-gold amalgamation method. J Hyg Chem 20:349-354 Weiss AA, Murphy SD, Silver S (1977) Mercury and organomercurial
- Weiss AA, Murphy SD, Silver S (1977) Mercury and organomercurial resistances determined by plasmids in *Staphylococcus aureus*. J Bacteriol 132:197-208
- Received February 8, 1988; accepted May 11, 1988.